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Association of AM fungi in Ricinus communis plants of Osmanabad

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ARTICLEINFO	ABSTRACT				
<i>Keywords:</i> AM fungi <i>Ricinus communis</i> Root colonization	A survey of the arbuscular mycorrhizal (AM) status associated with <i>Ricinus communis</i> plants growing and distributed in Osmanabad district of Maharashtra state was conducted. The result showed that all the different sites <i>R. communis</i> plants had AM fungal association in the roots and spore population in the rbigographic coil. However, maximum percent root colonization of AM fungi was observed in Parando sites				
*Corresponding author. E-mail addresses: ppsarwade@gmail.co m	(98 %) followed by others, while minimum in Tuljapur (62%). Paranda sites (345) showed more spore density whereas less in Kallam sites (104). Total five genera of AMF were identified in which <i>Acaulospora spp</i> and <i>Glomus spp</i> were found dominate followed by <i>Sclerocystis spp</i> and <i>Gigaspora spp</i> were found poorly distributed.				

1. Introduction

Arbuscular mycorrhizal fungi (AMF) form a symbiotic association with majority of land plants improving plant growth. More than 80 percent of all plants are associated with AMF in their root system (Smith and Read, 1997). These wellestablished AMF contribute to the phosphorus nutrition of plants by enhancing phosphorus uptake from the soil (Draft and Nicolson, 1966). Ricinus communis L. (Castor oil plant) is an annual or perennial shrub belonging to the family Euphorbiaceae. Leaves have long petiole and palm like lobed blades. Inflorescence consists of unisexual flowers which are arranged at the top of the axis in the form of panicles; male flowers lie towards the base and female flowers towards the apex; perianth leaves (sepals and petals) are inconspicuous and caducous. Fruit is three chambered, globose capsule with soft spines (Jombo and Enenebeaku, 2007). The chemical constituents showed the presence of amino acids (Onwuliri and Anekwe, 2001), fatty acids (Salimon et al., 2010), flavonoids (Ramos-Lopaz et al., 2010), phenolic compounds (Singh et al., 2009), phytosterol (Zhang et al., 2007), terpenoids (Darmanin et al., 2009), and other compounds (Ross, 2003) such as alkaloids, etc (Jena and Gupta, 2012). R. communis exhibits various biological and pharmacological activities such as abortifacient effect, acid phosphatase inhibition, acid

phosphatase stimulation, agglutin activity, alkaline phosphatase inhibition (Ross, 2003), anticonceptive activity (Okwuasaba et al., 1997), antidiabetic activity (Shokeen, 2008), antifertility effects (Sandhyakumary et al., 2003), anti-inflammatory activity (Singh et al., 2009), antimicrobial activity (Garcia et al., 2009), antioxidant activity (Singh et al., 2009), free radical scavenging activity (Ilavarasan et al., 2006), hepatoprotective activity (Visen et al., 1992), Insecticidal activity (Upasani et al., 2003), and repellent Properties (Grant, 2012).

Hence a study survey was conducted around Osmanabad district in Marathwada region, where the plant is grown throughout the year to observe AM fungal genera and species that are associated with plants.

2. Materials and Methods

Rhizosphere soil and roots samples of *R. communis* plants were collected from different locations of Osmanabad district (Viz. Kallam, Omerga, Paranda, Osmanabad, Tuljapur, and Bhoom) from each plant with three replications. Root samples were cleared and stained using Phillips and Hayman (1970) technique. Root colonization was measured according to the Giovannetti and Mosse (1980) method. Hundred grams of rhizosphere soil samples were analyzed for their spore isolation by wet sieving and decanting method Gerdemann and Nicolson, (1963). Identification of AM fungal genera up to species level by

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Sr No.	Plant species	Colonization	Types of	*Spore	AM fungal genera
		(%)	colonization	population	
1	Kallam	80	Н	104	Glomus spp
					Acaulospora spp
2	Omerga	65	HV	245	Glomus spp
					Acaulospora spp
					<i>Gigaspora</i> spp
3	Paranda	98	HV	345	Glomus spp
					Acaulospora spp
					Sclerocystis spp,
					Entrophosphora spp
4	Osmanabad	74	HV	225	Glomus spp
					Entrophosphora spp
					Acaulospora spp
5	Tuljapur	62	Н	252	Glomus spp
					Acaulospora spp
6	Bhoom	67	HV	210	Glomus spp
					Acaulospora spp

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*Mean of three samples, H- Hyphae V- Vesicular

using the Manual for identification by Schenck and Perez (1990).

3. Results and Discussion

The result shows that, maximum percent of colonization were found in Paranda sites (98 %) than other five sites whereas, minimum percentage was found in Tuljapur (62%). Hyphal and vesicular types of colonization were found in roots of different R. communis plants. Maximum number of spores (345) was observed in rhizosphere soil of Paranda sites than Kallam, Omerga, Osmanabad, Tuljapur, and Bhoom sites. Total five genera were observed viz., Acaulospora spp Glomus spp, Sclerocystis spp, Entrophosphora spp and Gigaspora spp. Highest number of AMF genera was associated with Paranda sites while the less number of AM fungal genera were recorded in other five locations. Among AM fungal genera Acaulospora spp and Glomus spp were found dominate followed by Sclerocystis spp, Entrophosphora spp and Gigaspora spp were found poorly distributed. The data of percent of colonization and spore number associated with R. communis plants different Osmanabad sites are presented in Table 1.

The occurrence of AMF in plants has reported earlier by Udea et al., (1992), Muthukumar and Udaiyan (2001). Recently Muthukumar et al., (2006) and Prakash et al., (2012) reported the occurrence of AMF in different plants from India.

4. Conclusion

The highest number of mycorrhizal spores in rhizosphere soil and AM fungal infection in the roots of *R. communis* indicated that these plant species might be considered good

itesThe author declares that there are no conflicts of interest.%).**References**

Conflicts of Interest

Darmanin, S, Wismayer, P.S., Podesta, M.T.C., Micallef, M.J. and Buhagiar, J.A. 2009. Phytochemistry. An extract from *Ricinus communis* L. leaves possesses cytotoxic properties and induces apoptosis in SK-MEL-28 human melanoma cells. Natural Product Research, 23(6): 561-571.

host for AMF under natural conditions. Therefore, here

concluded that, occurrence or distribution of AMF varies with different Osmanabad sites associated with *R. communis* plants.

Draft, M.J. and Nicolson, T.H. 1966. The effect of endogone mycorrhizae on plant growth. New Phytologist. 65:343-350.

Garcia, LFR., Almeida, G.L., Fernanda, C.P., Souza, P.D. and Consani. S. 2009. Antimicrobial activity of a calcium hydroxide and Ricinus communis oil paste against microorganisms commonly found in endodontic infections. Chemical Business. 24(8): 50.

Gerdemann, J.W. and Nicolson, T.H. 1963. Spores of mycorrhizal *endogone* species extracted from soil by wet sieving and decanting. Trans. Br. Mycol. Soc.46:235-244.

Grant, B. Castor Beans & Ground Moles. Available at:http://www.gardenguides.com/131783-castor-beansground-moles.html Accessed on November, 5, (2012).

Ilavarasan, R., Mallika, M. and Venkataraman, S. 2006. Antiinflammatory and free radical scavenging activity of *Ricinus communis* root extract. J. Ethnopharmacol. 103(3):478-480. Jena, J. and Gupta, A.K. 2012. *Ricinus communis* Linn: A phytopharmacological Review. Inter. J. Pharm. Pharmaceut. Sci., 4(4): 25-29.

Jombo, G. and Enenebeaku, M. 2007. Antimicrobial susceptibility patterns of bacteria to seed extracts of Ricinus Communis: Findings of a preliminary study in Nigeria. The Internet J. Microbio. 4(1): 1-6.

Muthukumar, T. and Udaiyan, K. 2001. Vesicular arbuscular mycorrhizal association in medicinal plants of Maruthamalai Hills,Western Ghats,South ern India. J.Mycol. Pl. Pathol. 31(2):180-184.

Muthukumar, T., Senthilkumar, M., Rajangam, M. 2006. Arbuscular mycorrhizal morphology and dark septate fungal associations in medicinal and aromatic plants of Western Ghats, Southern India. *Mycorrhiza*. 17:11-24.

Okwuasaba, F.K., Das, S.C., Isichei, C.O., Ekwenchi, M. M., Onoruvwe, O. and Olayinka, A.O. et al. 1997. The anticonceptive and the effect on uterus of ether extract, 18312-J of *Ricinus communis*. Phytother. Res. 10: 97-100.

Onwuliri, V. A. and Anekwe, G. E. 2001. Amino Acids and other biochemical Components of *Ricinus communis* L. (Variety Minor), an anti-conceptive Seed. Pak. J. Biol. Sci. 4(7):866-868.

Phillips, J.M. and Hayman, D.S. 1970. Improved procedures for clearing root and staining parasitic and vesicular arbuscular mycorrhizal fungi for rapid assessment of infection. *Tans. Bri. Mycol. Soc.*55(1), 158-161.

Picman, J. and Picman, A.K. 1984. Auto toxicity in *Parthenium hysterophorus* L. and its possible role in control of germination. Biochem. Syst. Ecol. 12: 287-292.

Prakash, P. Sarwade., Kanade, M. B., Ambuse, M. G. and Bhale, U. N. 2012. Association of arbuscular Mycorrhizal Fungi in some angiospermic plants of Maharashtra, India. International Multidisciplinary Research Journal. 2(4):18-19.

Ramos-López, M. A., Pérez, G.S., Rodríguez-Hernández, P.C., Fefer, G. and Sánchez, M. 2010. Activity of *Ricinus communis* (Euphorbiaceae) against *Spodoptera frugiperda* (Lepidoptera: Noctuidae). Afri. J. Biotechnol. 9(9):1359-1365.

Ross, I. A. 2003. *Ricinus communis* L. *In*: Medicinal plants of the world: chemical constituents, traditional and modern medicinal uses. 2: 375-393.

Salimon, J, Noor, DAM., Nazrizawati, A. T., Firdaus, MYM. and Noraishah, A. 2010. Fatty acid composition and physicochemical properties of malaysian castor bean *Ricinus communis* L. Seed Oil. Sains Malaysiana. 39(5): 761-764.

Sandhyakumary, K., Bobby, R.G. and Indira, M. 2003. Antifertility effects of *Ricinus communis* (Linn) on rats. Phytother. Res., 17(5): 508-511. Schenck, N. C. and Perez, Y. 1990. Manual for the identification of vesicular arbuscular mycorrhizal fungi. Synergistic Publications:Gainesville, FL.,U.S.A.,1-286.

Shokeen, P., Anand, P., Murali, Y.K. and Tandon, V. 2008. Antidiabetic activity of 50% ethanolic extract of *Ricinus communis* and its purified fractions. Food and Chemical Toxicalogy, 46(11):3458-3466.

Singh, P.P., Ambika and Chauhan, SMS. 2009. Activity guided isolation of antioxidants from the leaves of *Ricinus communis* L. Food Chemistry, 114:1069-1072.

Smith, S.E. and Read, D.J. 1997. Mycorrhizal symbiosis, 2nd Ed. Academic, San Diego, CA.

Udea,T., Husope, T., Kubo, S. and Nakawashi, I. 1992. Vesicular arbuscular mycorrhizal fungi (Glomales) in Japan II. A field survey of vesicular arbuscular mycorrhizal association with medicinal plants in Japan. Trans. Br. Mycol. Soc. 33:77-86.

Upasani, S.M., Kotkar, H.M., Mendki, P. S. and Maheshwari, V. L. 2003. Partial characterization and insecticidal properties of *Ricinus communis* L. foliage flavonoids. Pest Manag Sci., 59(12):1349-1354.

Visen, P.K.S., Shukla, B., Patnaik, G. K., Tripathi, S.C., Kulshreshtha, D.K., Srimal, R. C. and Dhawan, B. N. 1992. Hepatoprotective activity of *Ricinus communis* leaves. Pharm. Biol. 30(4): 241-250.

Zhang, X., Han, F., Gao, P., Yu, D. and Liu, S. 2007. Bioassay guided fractionation of antifertility components of castorbean (*Ricinus communis* L.) seed extracts. Nat. Prod. Res., 21(11): 982-989.